



Danuphylline, a novel pentacyclic indole from *Kopsia*

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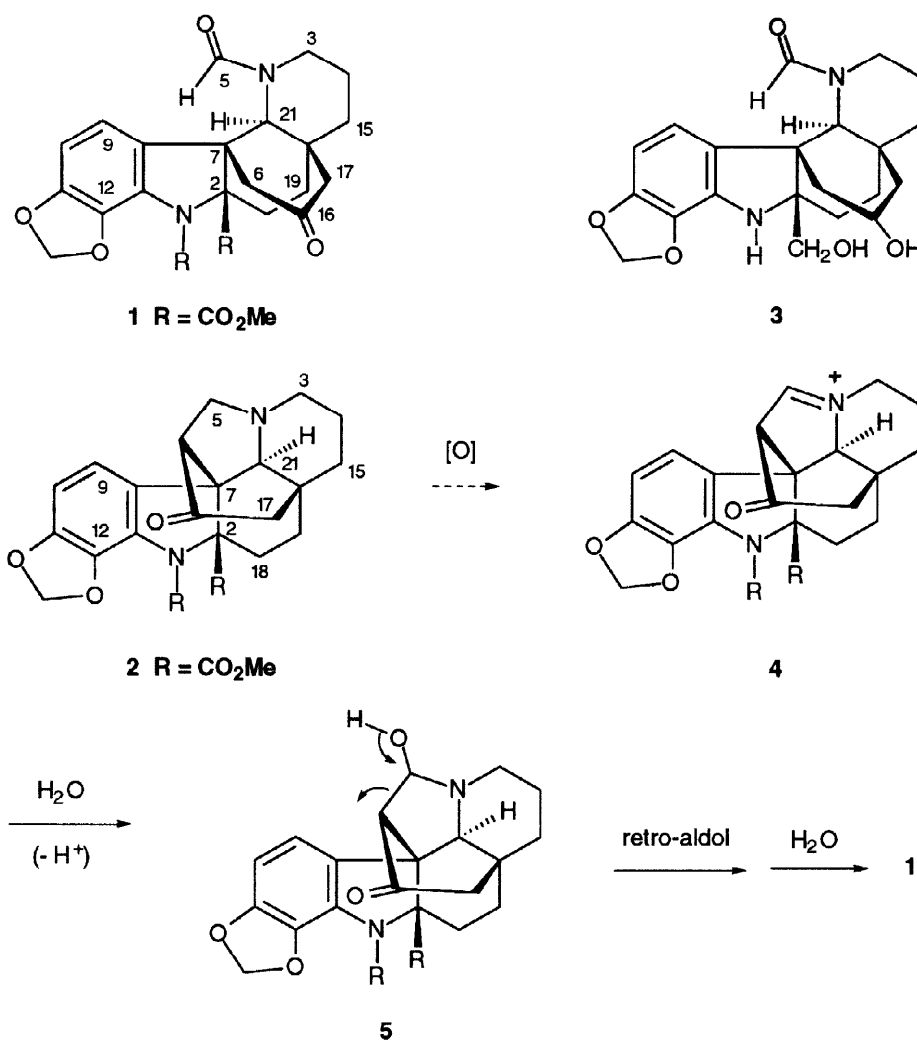
Abstract : A novel pentacyclic indole alkaloid, danuphylline, was obtained from the leaf-extract of *Kopsia dasyrachis* and its structure elucidated by spectral analysis.

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In the course of an investigation of the alkaloidal composition of the North Borneo species, *Kopsia dasyrachis*,¹ we obtained minor amounts of a new indole alkaloid, danuphylline, which possesses a novel pentacyclic carbon framework.

Danuphylline **1** was obtained in amorphous form, $[\alpha]_D -30^\circ$ (CHCl_3 , c 0.067). The UV spectrum showed absorption maxima at 214, 225, 248, 283 and 293 nm ($\log \epsilon$ 3.95, 4.13, 3.66, 3.03 and 2.99 respectively) typical of a dihydroindole chromophore, while the IR spectrum showed bands due to various carbonyl functions (1720 cm^{-1} , broad; 1669 cm^{-1}). The EIMS of **1** showed a molecular ion at m/z 470 with other significant fragment peaks at m/z 411 ($\text{M} - \text{CO}_2\text{Me}$, base) and 383 ($\text{M} - \text{CO}_2\text{Me} - \text{CH}_2=\text{CH}_2$). HREIMS measurements² gave the formula $\text{C}_{24}\text{H}_{26}\text{N}_2\text{O}_8$. The ^{13}C NMR spectrum showed a total of 24 carbon resonances, in agreement with the formula derived from the molecular ion. The ^1H and ^{13}C NMR spectral data (Table 1) showed the presence of a methylenedioxy substituent at carbon-11 and -12 (an AB doublet at δ_{H} 6.02, 5.97, J 1.5 Hz, δ_{C} 100.9), a CO_2Me substituent at N_1 (δ_{C} 153.4), a ketonic carbonyl (δ_{C} 206.7), an ester function (δ_{C} 170.2) and another carbonyl function associated with a formamide group (δ_{H} 6.68, δ_{C} 165.7; IR 1669 cm^{-1}).³ COSY and HMQC experiments revealed the presence of the following partial structures, viz., 2 isolated methylene groups, a $\text{CCH}_2\text{CH}_2\text{C}$ unit and a $\text{CCH}_2\text{CH}_2\text{CH}_2\text{N}$ fragment. The NMR data indicate that compound **1** possesses some of the features common to a basic aspidofractinine skeleton and is in some respects similar to that of the methylchanofrutosinate **2** (see Table 1) which is the major

alkaloid present in the leaves, and which has been obtained previously from various *Kopsia* species.^{4,5} There are however some notable differences. One major difference is the absence of signals due to the C(5)-C(6) unit and the appearance instead of an additional isolated methylene which is adjacent to a carbonyl function (δ_{H} 2.40, 2.74). That this methylene (C-6) is bridged by a ketonic function (C-16) to another isolated methylene (C-17) is indicated by the observation in the COSY spectrum of long range coupling (4J , W) from one of the H-6 to one of the H-17. Except for this long range coupling, and the expected geminal coupling, the hydrogens of both the C-6 and C-17 methylenes show no other coupling to any adjacent proton, providing confirmation that they are linked to quaternary centres. The direct attachment of the C-6 methylene to C-7 is supported by the correlation from C-7 to H-6 in the HMBC spectrum. Likewise, a similar correlation from



C-20 to H-17 confirmed the direct attachment of the C-17 methylene to the quaternary C-20. Another major difference shown by compound **1** when compared with **2** is the appearance of a formamide function (δ_{H} 6.68, δ_{C} 165.7),³ which from the HMBC data is deduced to be at position-5 (3J , C-5 to H-3, H-21). The location

of a formamide function at position-5 is also consistent with the unusual deshielding observed for H-3 β (δ_{H} 4.57) as a result of the anisotropy due to the proximate formamide carbonyl function, which has been observed previously in other compounds.⁶ These observations and other correlations from HMBC experiments (Table 1) suggest structure **1** for the new compound which represents a novel skeletal arrangement in which a new 6-membered ring incorporating C-7, -6, -16, -17, -20 and -21 has been formed by cleavage of the C-5/C-6 bond of the precursor compound **2**. Compound **1** can thus be considered a “*seco*-methylchanofruticosinate” and represents the first member of this novel group obtained as a natural product. Irradiation of the H-21 resonance causes NOE enhancement of the formamide resonance and *vice-versa*, indicating that the H-21 and the N-4 lone pair are now in a *trans* arrangement, in contrast to the conformation adopted in other aspidofractinine compounds such as kopsingine⁷ and the methylchanofruticosinate **2**.⁴

Table 1. ¹H and ¹³C NMR Spectral Data^a for **1** and **2**^b

Position	$\delta_{\text{C}}(2)$	$\delta_{\text{C}}(1)$	$\delta_{\text{H}}(1)$	² <i>J</i>	HMBC (1)	³ <i>J</i>
2	76.0	79.1	-	18		6, 19
3 α	46.5	34.7	2.70 m	14		5, 15, 21
3 β	-	-	4.57 dd (14.5, 9.5)	-		-
5	52.5	165.7	6.68 s	-		3, 21
6 α	55.5	39.3	2.40 br d (17)	-		17, 21
6 β	-	-	2.74 d (17)	-		-
7	58.4	54.5	-	6, 21		9, 18
8	129.1	125.0	-	9		6, 10, 21
9	116.9	117.0	6.35 d (8)	-		-
10	103.4	103.6	6.56 d (8)	-		-
11	149.0	150.4	-	10		9, OCH ₂ O
12	133.9	134.9	-	-		10, OCH ₂ O
13	124.1	124.9	-	-		9
14a	17.5	19.3	1.73 m	3, 15		-
14b	-	-	1.95 m	-		-
15a	35.3	29.6	1.29 dt (13.5, 9)	14		3, 17
15b	-	-	1.65 m	-		-
16	208.1	206.7	-	6, 17		-
17a	43.1	46.1	2.48 d (20)	-		6, 15, 19, 21
17b	-	-	2.72 d (20)	-		-
18a	23.5	23.0	2.36 ddd (16.5, 12, 8)	19		15
18b	-	-	3.28 dt (16.5, 3.5)	-		-
19a	34.9	39.4	1.65 m	18		17
19b	-	-	1.65 m	-		-
20	36.0	34.5	-	15, 17, 19, 21		-
21	68.5	61.5	3.38 s	-		3, 5, 6, 15, 19
OCH ₂ O	100.7	100.9	5.97 d (1.5); 6.02 d (1.5)	-		-
NCO ₂ Me	52.6	53.1	3.88 s	-		-
NCO ₂ Me	153.0	153.4	-	-		OMe
CO ₂ Me	53.0	53.0	3.63 s	-		-
CO ₂ Me	170.9	170.2	-	-		18, OMe

^aCDCl₃, 400 MHz; assignments based on COSY, HMQC, HMBC and NOE; ^b¹³C NMR data only

Examination of models indicates that there should be considerable hindrance to free rotation about the formamide C-N bond due to the proximity of the formamide function to the aromatic ring.⁸ Rotation about the C-N bond would result in severe repulsive interactions as the π -electron density of the formamide C=O as well as that of the 2 lone-pairs of the oxygen are brought into undue proximity with the π -electron density of the aromatic system. For this same reason a preferred conformation appears to have been adopted such that the formamide carbonyl is directed away from the aromatic ring, which in turn results in the formamide-H being placed within the shielding zone of the aromatic ring current, thus accounting for the unusually high field resonance observed for the formamide-H. Additional confirmation of the structure is provided by reduction ($\text{NaBH}_4/\text{MeOH}$, 3 h, 30 °C) of **1** which yielded a single product **3**. The hydroxymethyl protons of **3** are clearly seen as a pair of AB doublets (δ 3.21, 3.84; J 10.6 Hz) as is the H-16 oxymethine which is seen as a doublet of doublets at δ 4.32 (J 9, 7 Hz). The correlations observed from the hydroxymethyl protons to C-2 (2J) as well as to C-7 and C-18 (3J) provided confirmation for the attachment of the ester function at C-2 in danuphylline **1**. Furthermore, the three bond correlation from H-16 to C-7 and C-20 in the HMBC spectrum of **3** provided further confirmation for the location of the C-16 carbonyl function in danuphylline **1**.

A possible origin of this ring-opened alkaloid^{3,9} is from the methylchanofrucosinate **2** which on oxidation provides the iminium ion **4**. Hydrolysis of this iminium ion **4** gives, the presumably unstable carbinolamine **5**, which could then undergo a retro-aldol-type reaction to provide the *seco*-compound, danuphylline **1**. We have succeeded in realising such a biomimetic conversion of **2** to **1** based on the above proposal via the electrochemically-generated iminium ion **3**¹⁰ which will be reported in a forthcoming submission.

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References and Notes

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